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Impact of Polishing on Physicochemical Properties, Cooking Behaviour, and Anthocyanin Content in *Njavara* Red Rice

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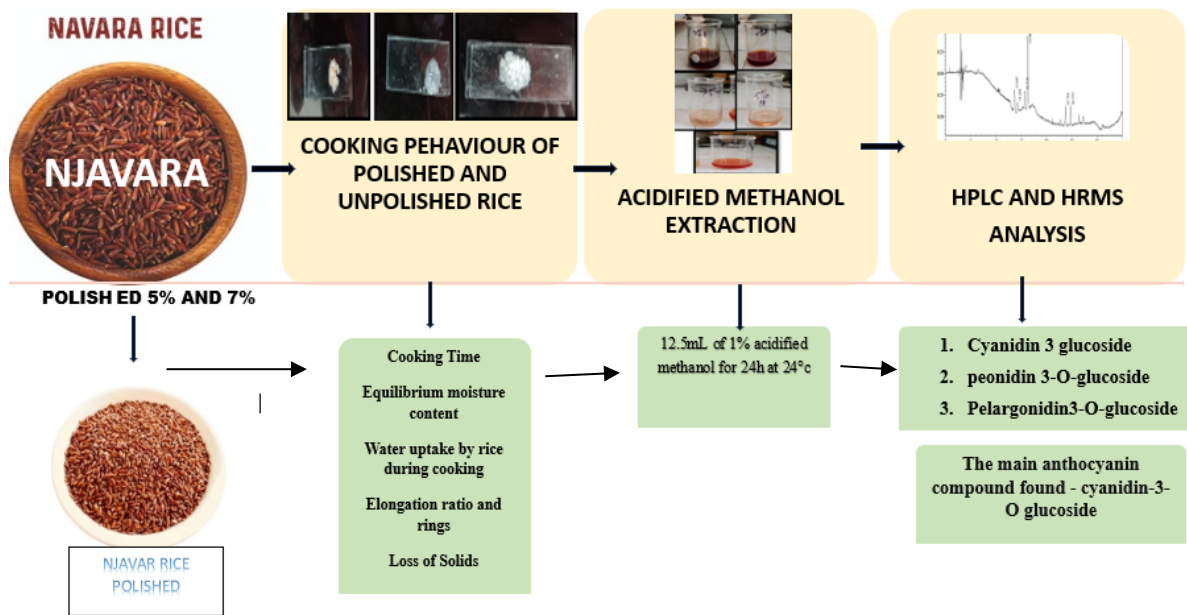
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Abstract

The aim of this study was to assess how rice polishing properties affect the nutritional value and anthocyanin composition and cooking performance of *Njavara* rice which grows as a traditional red rice variety in Kerala's Indian region. Red rice exists as unpolished rice which was examined together with 5% polished rice and 7% polished rice including their bran components. Red rice contains the highest nutritional value because it holds 8.565 mg GAE/g of total phenolic content which exceeds the 8.180 mg GAE/g found in 5% polished rice and 7.537 mg GAE/g found in 7% polished rice. Total anthocyanin content was highest in the 7% bran fraction (8.28 ± 0.06 mg/100 g), followed by red rice (6.81 ± 0.05 mg/100 g), 5% bran fraction (4.56 ± 0.08 mg/100 g), 5% polished rice (3.51 ± 0.02 mg/100 g), and 7% polished rice (2.53 ± 0.03 mg/100 g). The cooking properties showed red rice needed the longest cooking duration of 45 minutes while 5% polished rice needed 35 minutes and 7% polished rice needed 30 minutes with respective elongation ratios of 1.11, 1.18, and 1.21. The protein content was higher in red rice (9.42 ± 0.15%) as compared 9.23 ± 0.56% found in 5% polished rice and 8.12 ± 0.48% present in 7% polished rice. The research shows that polishing reduced bioactive compounds and antioxidant potential while decreasing cooking time and increasing solid loss during cooking.

Keywords: *Njavara*, Anthocyanin, Cooking Properties, Polishing, Nutritional Composition.



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33 1. Introduction

34 Rice (*Oryza sativa* L.) is an essential crop for more than half of the world's population and accounts for
 35 over 20% of the total calorie intake in the world with 90% of the total global production from Asia [1].
 36 White rice is the most consumed rice variety throughout the planet. Some Asian countries prefer the
 37 colored varieties such as red, browns, black, reddish-brown, and purple-black rice.

38 Pigmented rice, especially the Njavara variety, has great significance in tropical areas like
 39 Kerala, India, which is famous for its varied cultivars and the healing properties it has in Ayurveda. The
 40 Njavara rice, which is characterized by its white or black husks, has been the medicine of choice by
 41 Indian folk healers for ages. The dehusked Njavara rice is made up of 73% carbohydrates, 9.5% protein,
 42 2.5% fat, 1.4% ash, and provides 1628 kJ per 100 g of energy [2]. On the other hand, milling can take
 43 away much of the rice's nutrients, as the different nutrients (protein, fat, fiber, minerals, oryzanol,
 44 thiamine, and phenolic compounds) which the embryo and bran layer that are removed are concentrated
 45 in, play a crucial role in controlling rice quality characteristics [3].

46 Pigmented rice, which comprises the varieties with a bran layer that has not been peeled off
 47 and also colored pericarps such as black/purple and red, gives a rice of better nutritional value than
 48 white one [4]. The colors are influenced by the presence of acetylated procyanidin, anthocyanins, and

49 other phenolics that have considerable free radical scavenging activity [5], [6]. While pigmented rice
50 has been used mainly in desserts and special foods for festivals and religious offerings, it is now being
51 considered more and more as a delicacy and a source of nutrients with medicinal and pharmacological
52 potential[7]. Different types of rice have different colors with different anthocyanins, and the types and
53 amounts of anthocyanins depend on the rice variety. As rice processing goes on, the color of the rice
54 and the amount of anthocyanins decrease, which has been confirmed by the significant reduction of the
55 rice color during the milling process [8], [9], [10]. Kato & Horibata, (2021) [11] proved that the
56 antioxidant activity of polished rice is lower than that of red rice. Low milling degree (>4%) preserved
57 total anthocyanins in high amounts and the sensory quality of the sample was found to be good, but
58 total anthocyanins loss was in the range of 53-91% while the milling degree increased from 4% to 7%
59 [12]. Additionally, at the milling degree of 9% almost 100% of anthocyanins loss occurred[8]. This
60 showed that not only different rice milling degrees resulted in loss of nutrients but also that the
61 anthocyanins content was mainly confined in the bran of colored rice. Polishing preferentially removes
62 aleurone/bran layers, slashing total anthocyanins by 53-91% from 4-7% degree and nearly 100% at 9-
63 12%; profiles shift from diverse glycosides in red rice to trace monomers in polished forms. Njavara
64 specifically loses red pigmentation and antioxidant capacity, with bran fractions retaining highest levels
65 (e.g., 4-8x more than grains) [13]:

66 Furthermore, Lu et al. (2021) reported that rice's water absorption ability declined during
67 cooking, which might be due to the glutenin and starch interaction[14]. For a long time, people have
68 been using it in traditional medicine and as a functional food, but not much scientific literature has been
69 written about its response to different processes like these, which are important to the preservation of
70 its nutritional and functional qualities. Prior HPLC-HRMS work focused on generic black/red rice (2-
71 30 compounds), overlooking Njavara's unique Ayurvedic profile and precise low-degree polishing (5-
72 7%) impacts on cooking-nutrient synergies. This investigation uniquely quantifies polishing effects
73 (0%, 5%, 7%) on Njavara rice's nutrients, detailed anthocyanin profiles via HPLC-HRMS, and cooking
74 metrics (time, elongation). It addresses gaps in varietal-specific data for minimally processed forms.
75 Hence, this study proposal is to unveil the masked facts by involving the degrees of milling in the
76 discussion and analyzing in terms of their nutritional composition and cooking properties. The present
77 investigation further evaluates the effect of polishing on the anthocyanin profile of Njavara rice through
78 high-performance liquid chromatography (HPLC) and high-resolution mass spectrometry (HRMS).

79 **Materials and methods**

80 Njavara rice for the sample preparation were obtained from the local market of Mysore, India. Rice was
81 stored at 4°C in a cold storage room and packed in airtight bags to prevent any insect or pest
82 contamination. All chemicals used for analysis were of analytical grade unless otherwise specified and
83 were mostly sourced from Sigma.

84 **2.1 Sample preparation**

85 Njavara rice had undergone cleaning, dehusking through dehusker and polishing with 5-7% moisture
86 content in a Satake rice polisher. The polished fractions were (5% and 7%) and dried in pulverizer.
87 Then it was sieved through 60 mesh. The sieved sample along with bran samples (5% and 7%) were
88 collected in a zip lock bag. The samples were then stored at 4 degrees for further studies.

89 **2.2 Extraction of anthocyanin using acidified methanol**

90 Anthocyanins were extracted following the method described by Constantin (2022) [15] with slight
91 modifications. Finely ground rice bran and rice flour samples were used for extraction. The solvent
92 employed was acidified methanol, prepared as 1% (v/v) hydrochloric acid in methanol. A solid-to-
93 solvent ratio of 1 : 12.5 (w/v) was maintained, wherein 1 g of sample was mixed with 12.5 mL of the
94 extraction solvent. The mixture was kept under static extraction conditions at 24 °C for 24 h to facilitate
95 efficient solubilization of anthocyanins. After extraction, the mixture was centrifuged using an
96 Eppendorf centrifuge (model 5430) at 2000 × g for 15 min. The clear supernatant was carefully
97 collected. To ensure maximum recovery of anthocyanins, the extraction process was repeated three
98 times on the residual pellet under identical conditions. The supernatants from all extractions were
99 pooled and stored at 4 °C until further analysis.

100 **2.3 Proximate analysis**

101 The proximate composition of the Njavara rice variety was analyzed according to AOAC method (2016)
102 [16], including determination of moisture content through two-hour drying at 130°C, fat extraction
103 using a Soxhlet apparatus. Ash was determined using the method of incinerating the sample in a furnace
104 at high temperatures (around 550°C). Crude protein, calculated by multiplying total nitrogen by a factor
105 of 6.25, were measured using the Dumas method [17]. Total carbohydrates in Njavara rice were
106 determined by difference, calculated as 100 minus the sum of moisture, crude protein, total fat, and ash
107 percentages.

108 **2.4 Cooking properties**

109 **2.4.1 Cooking Time**

110 The minimum cooking time for milled rice was followed by using the Ranghino test [18]. 5 grams of
111 the sample was taken out and boiled in 20mL of preheated distilled water in a water bath at 90°C.
112 Minimum cooking time was determined by pressing the sample between two glass plates until no white
113 core remained.

114 **2.4.2 Equilibrium moisture content attained by rice upon soaking in water at ambient temperature** 115 **(EMC-S)**

116 In a 100 ml beaker a proper amount of rice (5-15 grams) was taken. The water was given the rice enough
117 time to absorb and reach the moisture content it can hold with the passing of time. After the soaking
118 period, the rice was taken out of the water and excess surface moisture was removed with a paper towel.
119 The rice was weighed one more time and the last weight was noted.

120 **2.4.3 Water Uptake and Loss of solids**

121 The rice was weighed and then rinsed with cold water to remove excess starch which would otherwise
122 cause stickiness during the cooking process. A careful drainage was done through a mesh. Then the
123 measured amount of water required for cooking, which usually follows the ratio of 1 part rice to 1.5 to
124 2 parts water, according to the desired texture and the specific rice type, was added. The rice was cooked
125 then dried to get rid of the absorbed water and solid loss can be calculated along with water gain.

126 2.4.4 Elongation ratio and rings

127 For measuring the length of a sample of cooked rice grains, a vernier caliper was applied.

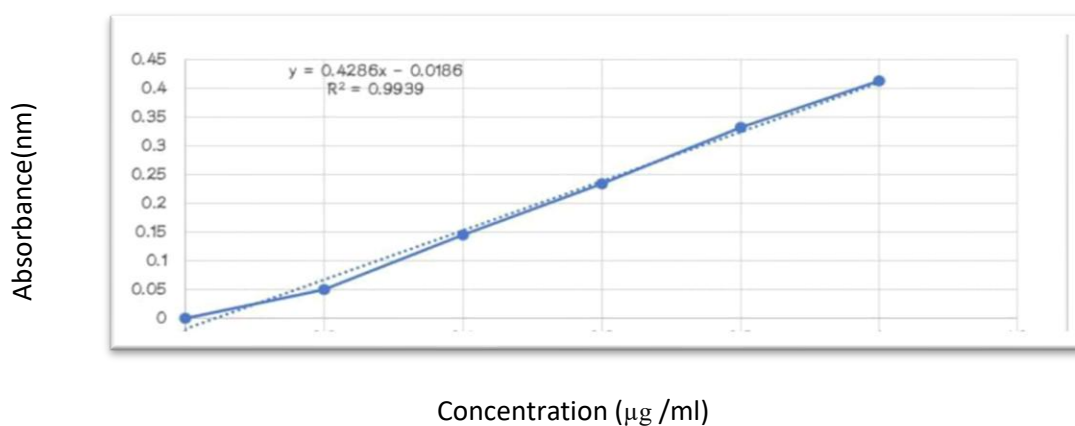
128 2.5 Bioactive compound analysis

129 2.5.1 Total Phenolic Content:

130 The total phenolic content of redand polished rice samples was determined using the Folin–Ciocalteu
131 (F–C) colorimetric method, following the procedure originally described by Martins et al., 2021 [19]
132 with appropriate modifications.

133 Briefly, the sample extract was diluted five-fold with distilled water. An aliquot of 0.2 mL of the diluted
134 extract was mixed with 6.0 mL of distilled water, followed by the addition of 0.5 mL of Folin–Ciocalteu
135 reagent. After 3 min of reaction, 1.5 mL of 20% (w/v) sodium carbonate (Na_2CO_3) solution was added,
136 and the final volume was adjusted to 10 mL with distilled water. The reaction mixture was incubated at
137 room temperature in the dark for 15 min, after which the absorbance was measured at 765 nm against
138 a reagent blank using a UV–Visible spectrophotometer . Gallic acid was used as the reference standard.
139 A standard calibration curve was prepared using different concentrations of gallic acid . The absorbance
140 values were plotted against gallic acid concentrations using Microsoft Excel (figure 1), and a linear
141 regression equation along with the coefficient of determination (R^2) was obtained [20].

142



149

150

151 Figure 1. Standard graph of Gallic acid

152

153 The total phenolic content was calculated from the calibration curve and expressed as mg gallic acid
154 equivalents per g of sample (mg GAE/g) using the corrected formula:

$$156 \text{ TPC (mg GAE/g)} = \frac{C \times V}{M}$$

155

157 where C is the concentration of phenolics obtained from the calibration curve (mg/mL), V is the volume
158 of extract (mL), and M is the mass of the sample (g).

159

160 **2.5.2 2,2-diphenyl-1-picrylhydrazyl (DPPH) Assay:**

161 40ul of sample extracts were mixed with 2.9ml DPPH- solution (4mg of DPPH in 100ml of methanol),
162 incubated in the dark at room temperature for 30 min, and absorbance measured at 517 nm [21] .

163 Inhibition calculated as % = $[(A_{\text{control}} - A_{\text{sample}})/A_{\text{control}}] \times 100$

164 **2.5.3 Determination of total Anthocyanin content**

165 The determination of total anthocyanin content, as described by Anggraeni et al., 2019 [22] involved
166 the following steps: The sample extract was mixed with two buffer solutions, maintaining a maximum
167 ratio of test portion to buffer at 1:4. The absorbance of the diluted test portion was then measured at
168 both 520 nm and 700 nm using buffers with pH 1.0 and pH 4.5. These measurements was taken against
169 a blank cell filled with distilled water. The absorbance measurements must be taken within 20-50
170 minutes of preparation to ensure accuracy in determining the total anthocyanin content.

171 **2.6 Differential Scanning Calorimetry (DSC)**

172 The instrument used was a PerkinElmer DSC8000 which was manufactured by PerkinElmer in U.S.
173 The rice sample (usually a few milligrams) was taken in a small aluminium pan. The sample pan and
174 the reference pan were placed in the DSC instrument. The experiment was started by heating or cooling
175 the pans at a controlled rate (usually constant) while recording the heat flow between the sample and
176 reference pans as a function of temperature.

177 **2.7 Scanning Electron Microscopy (SEM)**

178 Scanning Electron Microscopy (SEM) was done with the EVO® LS 15, a German built device
179 controlled by SmartSEM™ software. It was necessary to subject the sample to high-voltage electron
180 beam with energy from 100 to 30,000 electron volts. The image was created pixel by pixel as the
181 electron beam, which was directed by scan coils, moved in straight lines. The electron detector recorded
182 signals from the sample, such as secondary electrons (SE) and backscattered electrons (BSE). The SEM
183 delivered a partly 3D image which was influenced by the topography, i.e. the shape, size, and surface
184 texture, of the sample. The inclination of the sample surface not only influenced the topographic contrast
185 but also the number of BSE and SE signals, with an inclination of more than 50° to 70° boosting the
186 signals [23] .

187 **2.8 Quantification profile**

188 **2.8.1 HPLC- Instrumentation and chromatographic conditions.**

189 Anthocyanin content in rice bran and rice flour extracts was analyzed using a High-Performance Liquid
190 Chromatography (HPLC) system equipped with a gradient pump (Shimadzu LC-10AVp). The
191 chromatographic system was manufactured by Waters (Alliance series) and fitted with a UV-Visible
192 detector. Prior to HPLC analysis, the pooled methanolic extracts were filtered through Whatman No. 2
193 filter paper, transferred into amber vials, and stored at 4 °C until analysis. Chromatographic separation

194 was achieved using a Kromasil C18 reversed-phase column (particle size 5 μm , pore size 100 \AA ,
195 dimensions 4.6 mm \times 250 mm). The column temperature was maintained at 60 $^{\circ}\text{C}$. The injection volume
196 was 10 μL , and the flow rate was set at 1.0 mL min^{-1} . The mobile phase consisted of:

197 Solvent A: 0.1% (v/v) formic acid in water

198 Solvent B: 100% acetonitrile (HPLC grade)

199 A linear gradient elution program was employed as follows (Table 1):

200

201 **Table 1: Gradient elution program for HPLC analysis using Solvent A and Solvent B**

202

Time (min)	Solvent A (%)	Solvent B (%)
0	95	5
10	85	15
20	70	30
30	50	50
35	95	5
40	95	5

203 Detection of anthocyanins was carried out using a UV–Vis detector at 525 nm, which corresponds to
204 the maximum absorbance of anthocyanin pigments.

205 **2.8.2 High-Resolution Mass Spectrometry (HRMS)**

206 Further characterization and confirmation of anthocyanin compounds were performed using High-
207 Resolution Mass Spectrometry (HRMS). The analysis was carried out on a Sciex QTRAP 6500 /
208 TripleTOF 5600 mass spectrometer (Sciex, USA). Samples were ionized using both electrospray
209 ionization (ESI) and atmospheric pressure chemical ionization (APCI) sources. The instrument was
210 operated in both positive and negative ionization modes. The generated ions were introduced into a
211 hybrid quadrupole–time-of-flight (Q-TOF) mass analyzer, where separation occurred based on their
212 mass-to-charge (m/z) ratios. Data acquisition and processing were conducted using Analyst TF software
213 (Version 1.6). Mass spectra recorded predominantly in positive ion mode provided detailed information
214 on ion abundance and molecular mass, facilitating accurate identification of anthocyanin compounds
215 present in the samples.

216 **3 Results**

217 **3.1 Proximate analysis of Njavara rice**

218 The proximate composition of Njavara rice and its polished variants (5% polished, 7% polished, and
219 red rice) showed noticeable differences in moisture, fat, ash, and protein contents due to varying degrees
220 of polishing. The results are presented in Table 1. Moisture content ranged from $10.8 \pm 0.68\%$ in 5%
221 polished rice to $11.8 \pm 0.46\%$ in red rice, indicating a gradual increase in moisture retention with
222 reduced polishing intensity. Fat content was highest in red rice ($2.30 \pm 0.22\%$), followed by 5% polished
223 rice ($1.30 \pm 0.26\%$) and 7% polished rice ($1.12 \pm 0.24\%$), highlighting the contribution of bran layers
224 to lipid retention. Similarly, protein content was higher in red rice ($9.42 \pm 0.15\%$) and 5% polished rice
225 ($9.23 \pm 0.56\%$) compared to 7% polished rice ($8.12 \pm 0.48\%$). Ash content, which reflects mineral
226 concentration, also increased with lower polishing intensity, ranging from $1.09 \pm 0.58\%$ in 7% polished
227 rice to $1.74 \pm 0.72\%$ in red rice. These findings indicate that red rice retained superior nutritional quality,
228 while increased polishing resulted in progressive nutrient loss.

229 **3.2 Cooking Properties of Rice**

230 Cooking behavior varied significantly among the rice samples (Table 2). Red rice required the longest
 231 cooking time (45 ± 1.25 min), whereas 7% polished rice exhibited the shortest cooking time (30 ± 1.23
 232 min), followed by 5% polished rice (35 ± 1.26 min). The elongation ratio ranged from 1.11 ± 0.43 in
 233 red rice to 1.21 ± 0.25 in 7% polished rice, while 5% polished rice recorded an intermediate value of
 234 1.18 ± 0.74 . Equilibrium moisture content under normal conditions ranged from $10.2 \pm 1.23\%$ to 11.2
 235 $\pm 1.42\%$, and under soaked conditions from $11.2 \pm 1.52\%$ to $12.1 \pm 1.65\%$, with red rice showing
 236 comparatively higher moisture absorption. Water uptake was highest in 7% polished rice ($249.2 \pm$
 237 1.48%), followed by 5% polished rice ($241.7 \pm 1.56\%$), while red rice exhibited the lowest value (211.4
 238 $\pm 1.79\%$). In contrast, solid loss during cooking was highest in 7% polished rice ($7.2 \pm 1.56\%$) and
 239 lowest in red rice ($2.5 \pm 1.26\%$). These findings suggest that increased polishing reduced cooking time
 240 and enhanced water absorption and elongation, but also increased solid loss during cooking.

241 3.3 Bioactive Compound Analysis

242 The bioactive compound analysis presented in Table 2 revealed only slight variations in total phenolic
 243 content (TPC) among the samples. Red rice showed the highest TPC (8.565 ± 1.62 mg GAE/g),
 244 followed by 5% polished rice (8.180 ± 0.42 mg GAE/g) and 7% polished rice (7.537 ± 1.80 mg GAE/g).
 245 However, antioxidant activity measured by DPPH and ABTS assays differed more prominently among
 246 the treatments. Red rice exhibited the highest antioxidant activity, with DPPH and ABTS scavenging
 247 activities of $95.2 \pm 1.26\%$ and $97.6 \pm 0.52\%$, respectively. In comparison, 5% polished rice showed
 248 values of $90.5 \pm 1.23\%$ (DPPH) and $92.3 \pm 1.53\%$ (ABTS), while 7% polished rice recorded $89.7 \pm$
 249 1.52% and $91.5 \pm 1.24\%$, respectively. These findings indicate that reduced polishing helped preserve
 250 phenolic compounds and antioxidant potential, particularly in red rice.

251 3.4 Differential Scanning Calorimetry

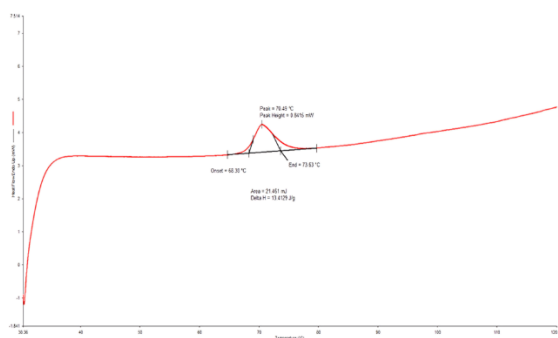
252 The differential scanning calorimetry (DSC) results presented in Table 1 and Figure 2a–c demonstrated
 253 variations in starch gelatinization behavior among the rice samples. Red rice exhibited the lowest onset
 254 temperature (62.81 °C) and peak temperature (66.53 °C), whereas 7% polished rice showed the highest
 255 peak temperature (70.85 °C) and end temperature (74.17 °C). The onset temperature of 5% polished
 256 rice was 68.30 °C, with a peak temperature of 70.49 °C and an end temperature of 73.63 °C. The higher
 257 gelatinization temperatures observed in polished rice samples may be attributed to the removal of bran
 258 layers and pigments, which exposed the crystalline starch structure and increased the thermal energy
 259 required for starch gelatinization.

260 **Table 2: Physico-chemical composition, thermal and cooking Properties of *Njavara* rice**

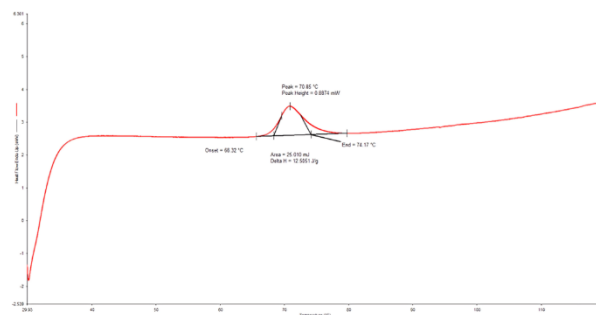
Analysis	5% polished rice	7% polished rice	Red rice
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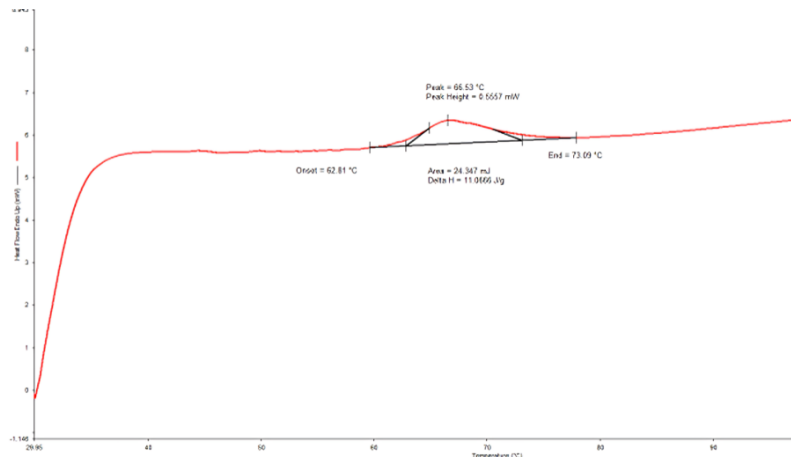
Differential scanning calorimetry			
Onset temperature (°C)	68.30	68.32	62.81
Peak temperature (°C)	70.49	70.85	66.53
End temperature (°C)	73..63	74.17	73.09
Proximate Analysis			
Moisture (%)	10.8±0.68 ^a	11.2±0.35 ^a	11.8±0.46 ^a
Ash (%)	1.30±0.25 ^a	1.09±0.58 ^a	1.74±0.72 ^a
Fat (%)	1.3±0.26 ^a	1.12±0.24 ^a	2.3±0.22 ^b
Protein (%)	9.23±0.56 ^a	8.12±0.48 ^b	9.42±0.15 ^a
Cooking Properties			
Cooking time(min)	35±1.26 ^b	30±1.23 ^a	45±1.25 ^c
Elongation ratio	1.18±0.74 ^a	1.21±0.25 ^a	1.11±0.43 ^a
Equilibrium moisture			
Normal	10.7±1.19 ^a	10.2±1.23 ^a	11.2±1.42 ^a
Soaked	11.5± 1.25 ^a	11.2±1.52 ^a	12.1±1.65 ^a
Water Uptake	241.7±1.56 ^b	249.2±1.48 ^a	211.4±1.79 ^c
Loss of solid (%)	6.2±1.43 ^b	7.2±1.56 ^a	2.5±1.26 ^c
Antioxidant profile			
Total Phenolic Content(mg GAE/g)	8.180±0.42 ^a	7.537±1.80 ^a	8.565±1.62 ^a
DPPH (%)	90.5±1.23 ^b	89.7±1.52 ^b	95.2±1.26 ^a
ABTS (%)	92.3±1.53 ^b	91.5±1.24 ^b	97.6±0.52 ^a

261



262





263

264

265 **Figure 2. DSC thermogram showing starch gelatinization characteristics of red rice a) 5% Polished rice**
 266 **b) 7% Polished rice c) Red rice**

267 **3.5 Total Anthocyanin Content**

268 Significantly lower amounts of anthocyanins were detected upon increasingly polished rice samples (Table 3).
 269 The maximum total anthocyanin content (TAC) was characterized in the 7% bran fraction (8.28 ± 0.06 mg
 270 C3G/100 g), then red rice (6.81 ± 0.05 mg C3G/100 g) and 5% bran fraction (4.56 ± 0.08 mg C3G/100 g)
 271 followed. The polished samples were a lot lower than this, with the 5% polished rice containing 3.51 ± 0.02 mg
 272 C3G/100 g and the 7% polished rice having only 2.53 ± 0.03 mg C3G/100 g.

273

Table 3: Total anthocyanin Content

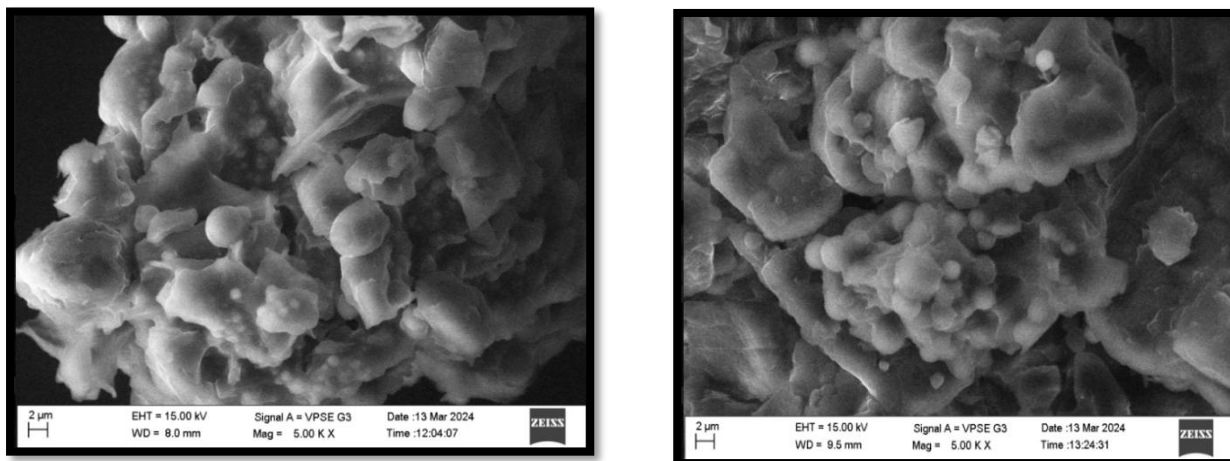
	5% bran fraction	5% polished rice	7% bran fraction	7% polished rice	Red rice
Total Anthocyanin Content (TAC) (mg/100g)	4.56 ± 0.08^c	3.51 ± 0.02^b	8.28 ± 0.06^c	2.53 ± 0.03^a	6.81 ± 0.05^d

274

275 **3.6 Surface morphology**

276 Red rice, due to its morphological properties, has a rough and grainy surface, and this uneven texture can be
 277 regarded as a sign of the original bran and germ layers. The intricate composition of the bran is not only revealed
 278 but also reflected in the variety of particle shapes and sizes on its surface, which is the result of the existence of
 279 compounds rich in anthocyanin. The keeping of the surface structure in red rice is very important for the retention
 280 of anthocyanins, which in turn helps the grain acquire its antioxidant properties and the health benefits associated
 281 with it, as indicated in Figure 3a . Smooth and uniform surfaces with the least irregularities show the characteristics
 282 of polished rice. The process of polishing, in which the outer bran and germ layers are removed, effectively
 283 destroys the main reservoirs of anthocyanins in the grain. Moreover, the presence of pores, voids, and structural

284 defects like cracks and fissures is related to the mechanical stress that the polishing process imposed. The particles
 285 of bran and germ that sometimes can be seen on the polished rice's surface are the traces of the anthocyanin-rich
 286 layers that were lost during the milling process, as shown in Figure 3b. Although the smoother appearance of
 287 polished rice has become the most common choice in cooking, the nutritional advantages of red rice, especially
 288 those coming from its anthocyanin content, are likely to draw the attention of health-conscious consumers.



289

290

291 **Figure 3: Scanning Electron microscopy of Rice a) Whole Rice Grain Surface b) Polished rice**

292

293 **3.7 Analysis and Quantification of Anthocyanin by HPLC**

294 The results from HPLC analysis conducted on Njavara rice samples indicated that cyanidin-3-glucoside was the
 295 major anthocyanin compound as seen in Table 4. The highest concentration in red rice (418.6 $\mu\text{g}/100\text{ g}$, Peak 6,
 296 retention time 13.885 min, peak area 36,965) was revealed by the quantification. The 5% bran fraction had 364.4
 297 $\mu\text{g}/100\text{ g}$ (Peak 10, retention time 13.199 min, peak area 32,183), while the 7% bran fraction was reduced to 191.7
 298 $\mu\text{g}/100\text{ g}$ (Peak 3, retention time 13.786 min, peak area 16,930). The content of cyanidin-3-glucoside in white rice
 299 was very low with the 5% white rice having 34.9 $\mu\text{g}/100\text{ g}$ (Peak 5, retention time 13.647 min, peak area 3,082)
 300 and the 7% white rice containing 3.5 $\mu\text{g}/100\text{ g}$ which was the lowest (Peak 13, retention time 13.656 min, peak
 301 area 305). The results of the study show that cyanidin-3-glucoside gets lost progressively and in large amounts
 302 when polishing is done, thus it means that most of this anthocyanin is present in the outer bran layers which get
 303 removed during the polishing process. They are represented in supplementary material as figures (Fig. S1-Fig S5).

304

Table 4: HPLC Peaks

Sample	Peak	Ret. Time	Area	Compound Name	Concentration $\mu\text{g}/100\text{g}$
5% Bran fraction	Peak 10	13.199	32183	Cyanidin 3 glucoside	364.4

7% Bran fraction	Peak 3	13.786	16930	Cyanidin 3 glucoside	191.7
5% Polished Rice	Peak 5	13.647	3082	Cyanidin 3 glucoside	34.9
7% Polished rice	Peak 13	13.656	305	Cyanidin 3 Glucoside	3.5
Red rice	Peak 6	13.885	36965	Cyanidin 3 glucoside	418.6

305

306 3.8 Identification and Characterization by High-Resolution Mass Spectrometry (HRMS)

307 The High-Resolution Mass Spectrometry (HRMS) analysis of *Njavara* rice samples confirmed the presence of
 308 key anthocyanin compounds based on their observed mass-to-charge ratios (m/z), which closely matched
 309 theoretical values. Cyanidin 3-glucoside was identified with an observed m/z of 287.05, corresponding exactly to
 310 its theoretical m/z of 287 and molecular formula $C_{21}H_{21}ClO_{11}$. Pelargonidin 3-O-glucoside appeared with an
 311 observed m/z of 271.05, matching the theoretical m/z of 271 ($C_{21}H_{21}ClO_{10}$). Peonidin 3-O-glucoside was detected
 312 at an observed m/z of 301, in agreement with the theoretical m/z of 301 and molecular formula $C_{22}H_{23}O_{11}Cl$ (Table
 313 5). These spectral data confirm that cyanidin, pelargonidin, and peonidin derivatives constitute the major
 314 anthocyanins in *Njavara* rice, supporting its characteristic pigmentation and antioxidant properties reported in the
 315 literature. The precise mass accuracy and characteristic molecular ions validate the molecular identity of these
 316 bioactive flavonoid compounds contributing to the rice's functional and therapeutic attributes. They are
 317 represented in supplementary material as figures (Fig. S6-Fig S8).

318

Table 5: Spectral Data

Compound	Observed m/z	Theoretical m/z	Formula
Cyanidin 3 glucoside	287.05	287	$C_{21}H_{21}ClO_{11}$
Pelargonidin 3-O-glucoside	271.05	271	$C_{21}H_{21}ClO_{10}$
peonidin 3-O-glucoside	301.07	301	$C_{22}H_{23}O_{11}Cl$

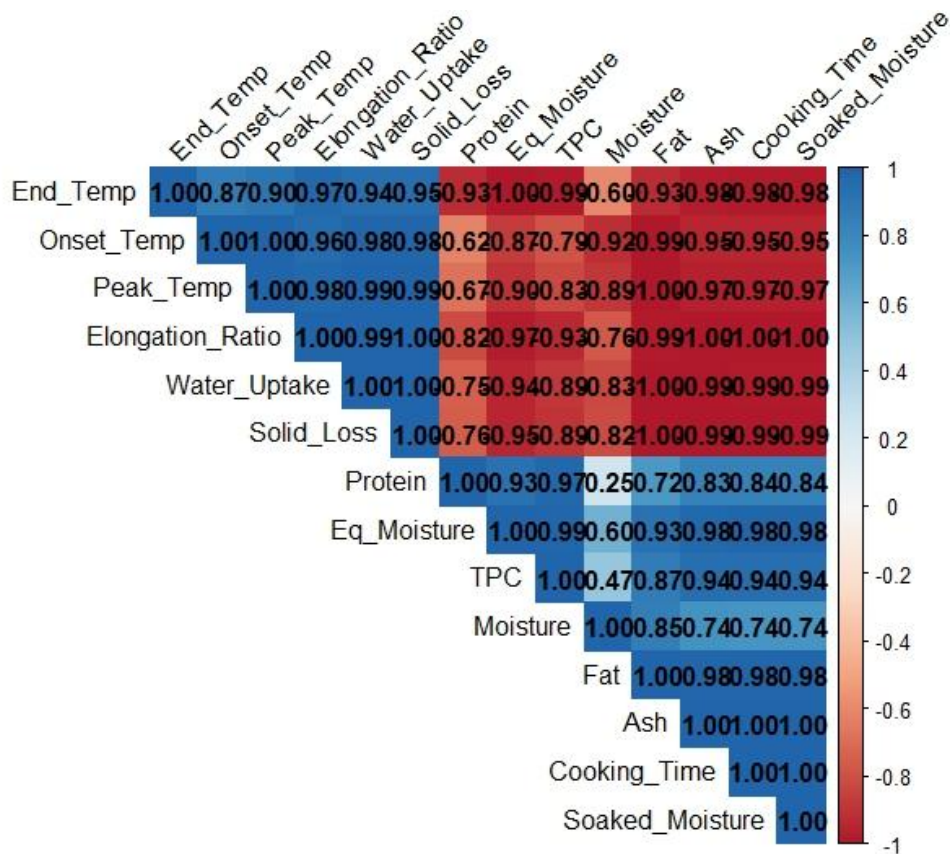
319

320 3.9 Correlation analysis

321 The correlation analysis of *Njavara* rice properties has shown some significant interrelationships between the
 322 thermal, cooking, nutritional, and bioactive parameters. The Pearson correlation matrix revealed that there were
 323 high positive correlations among thermal transition parameters (end temperature, onset temperature, peak

324 temperature), elongation ratio, water uptake, solid loss, protein content, equilibrium moisture, fat, ash, cooking
 325 time, and soaked moisture, with most correlation coefficients above 0.90. This points to a close relationship
 326 between these characteristics and their tendency to either increase or decrease together, thus demonstrating their
 327 interdependence in the grain matrix.. For instance, higher gelatinization temperatures correlated with improved
 328 grain expansion during cooking and prolonged cooking times, indicating that starch properties, protein content,
 329 and moisture uptake all play a substantial role in determining functional rice quality (Figure 4).

330 In contrast, the simplified matrix focusing on polishing, total phenolic content (TPC), and anthocyanin
 331 levels revealed strong negative correlations between polishing and both TPC ($r = -0.93$) and anthocyanin ($r = -$
 332 1.00). This result demonstrates that increasing the degree of polishing leads to significant depletion of antioxidant
 333 phenolics and anthocyanin pigments, which are vital for the rice's nutraceutical value. Furthermore, TPC and
 334 anthocyanin content showed a strong positive correlation ($r = 0.90$), indicating that anthocyanins make a major
 335 contribution to the overall phenolic profile and antioxidant activity of *Njavara* rice. Collectively, these results
 336 emphasize the crucial importance of minimal polishing for preserving nutritional, functional, and health-promoting
 337 properties of this traditional rice variety (Figure 5).



338
 339 **Figure 4: Correlation Matrix of Thermal, Cooking, and Nutritional Properties**

340

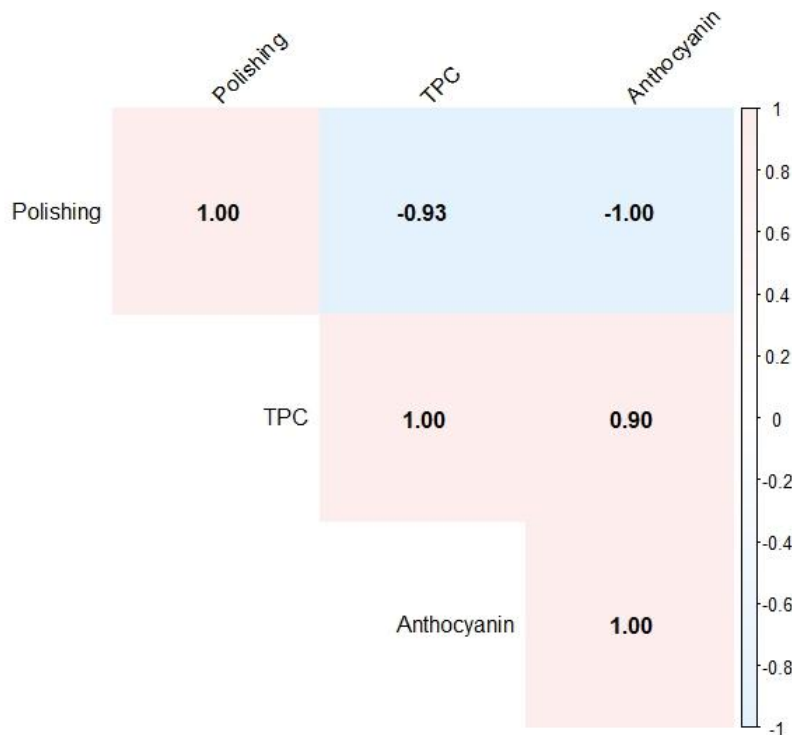


Figure 5: Correlation Matrix of Polishing Level with TPC and Anthocyanin Content

341

342

343

344 Discussion

345 The results of this research quite evidently indicate that the extent of milling has a great impact on the
 346 nutritional and functional properties of Njavara rice, which is in line with the previously observed trends for
 347 pigmented rice varieties. When all the grains were kept brown, they still managed to keep up to ten times their
 348 original amount of protein, fat, and mineral matter as well as moisture compared to 5% and 7% polished rice thus,
 349 confirming earlier studies [10], [24], [25]. These investigations had pointed out that the bran layers are the principal
 350 sources of the important nutrients and were confirmed by the current study which recorded fat and protein loss
 351 with the increase in the degree of polishing. The higher ash content, which is an indicator of the mineral richness,
 352 has backed the Deepa et al., 2008 [2] finding that the Njavara rice and pigmented genotypes, in general, are high
 353 in potassium, magnesium, iron, and phosphorus in the outer layers. As far as the cooking behavior is concerned,
 354 the decrease in cooking time with the rise of polishing is in line with the findings of Mardiah et al., (2017) [26],
 355 Nayeem et al., (2021) [27] who concluded that the reason for the faster cooking up was that the thinning of the
 356 husk allows the water to get in easily which in turn speeds up the gelatinization process. However, this is
 357 accompanied by a higher solid loss and a decrease in textural integrity that is in line with the increased leaching
 358 of more polished rice samples[10]. The acceptance of and improvement in texture after moderate (5%) polishing
 359 treatment, as seen in the present study, was similar to the findings of Mardiah et al., (2017) [26] who stated that a
 360 single mild polishing step not only conferred preferred sensory attributes on red rice without substantial sacrifices
 361 in nutritional quality but also led to developing consumer-preferred sensory attributes in red rice without major
 362 sacrifices in nutritional quality [26].

363 The current research outcomes also support the notion that Njavara rice is naturally packed with bioactive
 364 compounds. The measured total phenolic content (TPC) showed a very slight decrease from red to 5% polished
 365 rice, while in 7% polished samples the TPC was considerably lower, thus very close to the trends of the reports by

366 Priyanthi & Sivakanesan, 2021, Avinash et al., 2024) [28], [29] who claimed that phenolics localized in the bran
367 were the first to go during milling. The high antioxidant activities measured in red rice samples, which were
368 evaluated using DPPH and ABTS methods, are in agreement with the findings of Rai et al., (2025) [30] and Ed
369 Nignpense et al., (2024) [31], who reported that colored and traditional rice varieties always win over white and
370 fully polished rice in terms of antioxidant capacity thanks to their phenolic and flavonoid content. A very
371 remarkable result was the significant decrease of anthocyanins with polishing [30], [32], [33]. These researchers
372 contend that anthocyanins, mainly cyanidin-3-O-glucoside in red rice, are encompassed in the outer pericarp and
373 aleurone layers and hence very much prone to removal by even little polishing. The current results showing that
374 there was nearly total loss of anthocyanins after 7% polishing, strengthen the claim made by Priyanthi &
375 Sivakanesan, (2021) [28] that the unpolished grains may have up to 80% more anthocyanins than the polished
376 ones. The correlation data showing an inverse relationship between polishing and anthocyanin content ($r = -1.00$)
377 very precisely supports these literature conclusions. The thermal and textural changes due to polishing, which were
378 evidenced by DSC and microscopy in this research, are also highly cited in the literature.

379 Thermal and textural alterations following polishing, as demonstrated through DSC and microscopy in
380 this study, are similarly well documented in the literature. The observed upward shift in gelatinization temperatures
381 with increased polishing is explained by the removal of phenolic–lipid–amylose complexes, as discussed by [34],
382 and matches the reduction in lipid content postulated by Bani et al., (2024) [10] In addition, studies by Abdel-Aal
383 et al. (2006) [35] and Pereira-Caro et al., (2013) [36] reveal that anthocyanins and other phenolics not only serve
384 as antioxidants but also modify starch behavior during cooking by interacting with amylose and amylopectin
385 chains; their removal therefore increases gelatinization temperatures and changes textural outcomes.

386 The present study also aligns with reports that that post-milling handling—such as powdering and storage under
387 suboptimal conditions increasing the loss of anthocyanins, further compromising nutritional value. In reviewing
388 the comparative literature on Indian pigmented rice [37] . Ed Nignpense et al., (2024) [31] and Deepa et al., (2008)
389 [2] positioned *Njavara* as a leader in phenolic content and antioxidant activity, providing a strong rationale for
390 recommending minimal processing.

391 Overall, these findings establish that the traditional wisdom behind the use of red and lightly polished *Njavara* rice
392 finds strong scientific support: minimal polishing preserves the grain’s nutritional superiority and functional
393 benefits, while more extensive polishing, though improving cooking characteristics, results in significant depletion
394 of the compounds responsible for its medicinal and health-promoting properties. These results underscore the
395 importance of optimizing post-harvest processing and cooking methods to ensure that pigmented rice varieties
396 deliver both sensory appeal and maximum nutraceutical value in modern diets, as also advocated by multiple recent
397 studies in the field.

398

399 **Conclusion**

400 The present study comprehensively demonstrates the profound influence of polishing on the nutritional, functional,
401 bioactive, thermal, and structural attributes of *Njavara* rice—a traditionally valued, pigmented medicinal variety.
402 Minimal processing (5% polishing) retained much of the intrinsic nutritional value of red rice while significantly
403 improving cooking and textural properties, suggesting a favourable compromise between consumer acceptability
404 and health benefits. Future research should explore long-term health impacts, clinical validations, and
405 bioavailability studies of the preserved bioactive to fully harness *Njavara*’s nutraceutical potential.

406

407 **Conflicts of Interest**

408 The authors confirm that they have no conflicts of interest with respect to the work described in this manuscript."

409 **Authors Contributions**

410 Anju Augustine- Investigation, resources, data curation,

411 Naomi Vincent - writing—original draft preparation, writing, visualization, review and editing,

412 Dr. Shruti Joshi – Conceptualization, supervision

413 All authors have read and agreed to the published version of the manuscript.

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425 **Availability of Data and Materials:**

426 Data supporting the results of this study are available upon request from the corresponding author.

427 **List of abbreviations**

428 1. **HPLC** – High-Performance Liquid Chromatography

429 2. **HRMS** – High-Resolution Mass Spectrometry

430 3. **AOAC** – Association of Official Analytical Chemists

431 4. **EMC-S** – Equilibrium Moisture Content (Sorption)

432 5. **TPC** – Total Phenolic Content

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438

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